

which in turn may be converted into cularine- or turkiyenine¹⁶-type alkaloids, depending upon the enzymatic systems available in the plant.

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(+)-Turkiyenine: An Unusual Extension of the Biogenetic Sequence for the Isoquinoline Alkaloids

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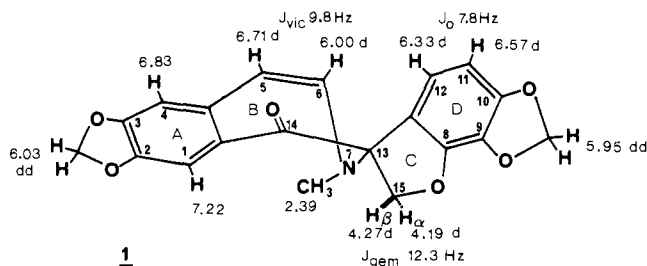
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The amorphous base turkiyenine (**1**) was originally isolated by us 2 years ago, shortly following collection of the plant *Hypecoum procumbens* L. (Papaveraceae) in April 1982, near the village of Fethiye, in the province of Muğla, in south central Anatolia. Our original studies indicated that the alkaloid C₂₀H₁₅NO₆ ($[\alpha]^{23}_D +72^\circ$ (*c* 0.053, CHCl₃); ν_{\max} CHCl₃ 1663, 1710 cm⁻¹; λ_{\max} MeOH 218 sh, 258, 281 sh, 352 nm (log ϵ 4.42, 4.62, 4.11, 3.55))¹ incorporated an *N*-methyl group, a ketonic function, a *cis*-disubstituted double bond as part of a seven-membered ring, and a spiro linkage—an amalgam of structural features hitherto unknown among the recognized isoquinoline- or isoquinoline-derived alkaloids.

In order to confirm the authenticity of (+)-turkiyenine (**1**) as an alkaloid, *H. procumbens* was collected again in 1983, on the same day and the same location where it had been gathered the previous year.² Extraction again provided (+)-turkiyenine, at

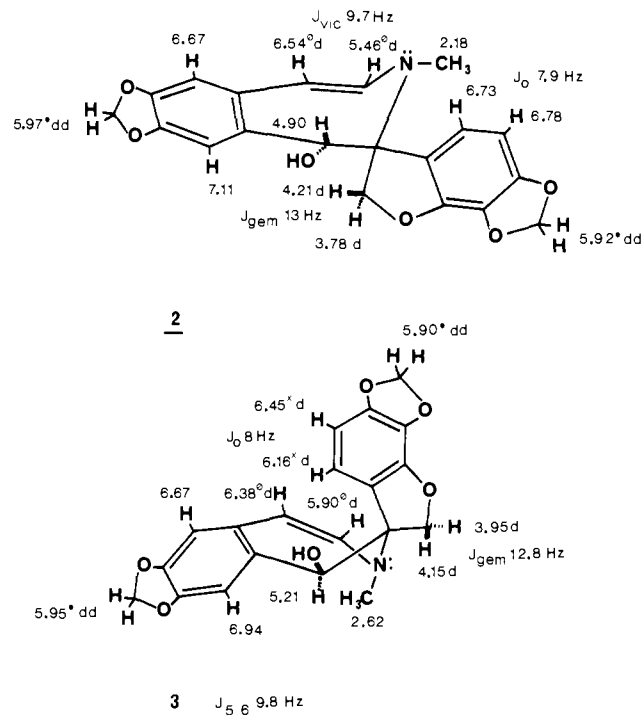


which point a detailed structural study became warranted.

The 360-MHz (CD₃CN) ¹H NMR spectrum of turkiyenine has been summarized around expression **1** of relative configuration.³ One *N*-methyl singlet is present at δ 2.39. Two methylenedioxy groups appear each as closely packed doublets of doublets, one set at δ 6.03 and the other at 5.95. The vinylic protons of the *cis* double bond are found at δ 6.00 and 6.71, each as a doublet with $J_{vic} = 9.8$ Hz. A particularly informative feature of the spectrum was the two-proton methylene doublets at δ 4.19 and 4.27, $J_{gem} = 12.3$ Hz. A coupling of this magnitude is characteristic of a five-membered ring containing an oxygen atom.⁴

NMR NOEDS⁵ proved to be particularly effective not only in the gross structural elucidation but also in the establishment of the favored conformation. The seven-membered ring is in a quasi-boat conformation. Irradiation of the *N*-methyl singlet (δ 2.39) caused enhancements of three signals, namely, H-1 (δ 7.22) by 3.0%, H-6 (δ 6.00) by 15%, and H-15 α,β (δ 4.19–4.27) by ~5%. Irradiation of the methylenedioxy protons at δ 6.03 resulted in a 2.6% NOE of the H-1 (δ 7.22) singlet which has a long T_1 since no other proton is situated in its immediate vicinity. It follows that the alternate methylenedioxy absorption at δ 5.95 can be assigned to the substituent on ring D. A significant negative NOE of 2.6% was recorded for H-4 (δ 6.83) upon irradiation of the H-6 signal (δ 6.00).⁶ This is a counterpoint to the strongly positive NOE of 19.4% observed for H-5 (δ 6.71) upon irradiation of H-6. Finally, by use of Gaussian multiplication, long-range coupling through five bonds could be detected between H-1 (δ 7.22) and H-5 (δ 6.71).

Reduction of (+)-turkiyenine (**1**) with sodium borohydride in methanol provided two amorphous alcohols, **2** and **3**, C₂₀H₁₇NO₆, in a 2:1 ratio. In both alcohols, the C-14 hydroxyl prefers to adopt



a pseudoequatorial position. For the major compound **2**, $[\alpha]^{23}_D -28^\circ$ (*c* 0.001, CHCl₃), two NMR NOE's were observed upon irradiation of the pseudoaxial H-14 at δ 4.90, viz., H-1 (δ 7.11)

(1) Turkiyenine: mass spectrum, m/z 364 ($M - 1$)⁺ (0.9), 363 (6), 362 (18), 350 (14), 349 (59), 332 (54), 320 (100); CD $\Delta\epsilon$ (nm) (MeOH) +1.2 (403), -9.5 (353), -2.4 (290), +22.4 (258), -10.1 (239).

(2) Plant collection dates were April 8, 1982 and 1983. The dried plant material (1.35 kg) was extracted with ethanol at room temperature. The usual acid-base workup, followed by silica gel chromatography, provided 30 mg of turkiyenine.

(3) We have found that CD₃CN is generally a superior solvent to CDCl₃ for recording the NMR spectra of alkaloids. The aromatic proton peaks are better separated, and the solvent is not as destructive of alkaloids as chloroform.

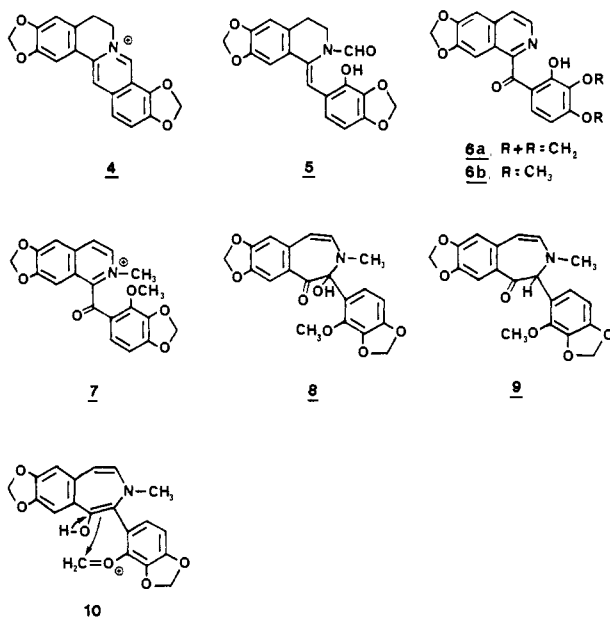
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by 2.9% and H-12 (δ 6.73) by 15.3%. The latter NOE was critical in confirming that the ring-D methylenedioxy was located at C-9,10 rather than at the alternate C-11,12 site. Turning now to the minor compound **3**, $[\alpha]_D^{23} +409^\circ$ (c 0.0007, CHCl_3), four NOE's were recorded following irradiation of the pseudoaxial H-14. The stronger two concerned the acidic C-14 alcoholic hydrogen at δ 3.12 (10.7%) and the *N*-methyl singlet at δ 2.62 (7.1%). Weaker effects were observed for the H-1 singlet at δ 6.94 (3.5%) and the H-15 β doublet at δ 4.15 (0.9%).⁷

The biogenesis of (+)-turkiyenine (**1**) appears to be radically different from that of any other isoquinoline-derived alkaloid. The origin of turkiyenine may hypothetically be traced to the pseudobenzylisoquinoline **5** which could be obtained biogenetically from the protoberberinium alkaloid coptisine (**4**).⁸ Hydrolysis and



oxidation of **5** would yield pseudobenzylisoquinoline **6a**, structurally related to the known rugosinone (**6b**).⁹ O,N-Dimethylation of **6a** would provide salt **7**, which through base-catalyzed isomerization could lead to pseudobase **8**. Reduction of **8** would give rise to ketone **9**. Enolization of this ketone, and oxidation of the aromatic methoxyl group to an oxonium ion as in **10**, would then set the stage for the formation of the methylenoxy bridge¹⁰ and the alkaloid (+)-turkiyenine (**1**).

It has been previously suggested that the intermediacy of pseudobenzylisoquinolines could be one of several possible ways by which 8,9,10-oxygenated aporphines may be formed in nature.⁸ It is now apparent that pseudobenzylisoquinolines may also be implicated in the biogenesis of the unusual base (+)-turkiyenine (**1**).

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(7) (-)-Dihydroturkiyenine (**2**): ν_{max} (CHCl_3) 3350 cm^{-1} ; λ_{max} MeOH 223, 288, 309 sh, 319 nm ($\log \epsilon$ 4.47, 3.92, 3.83, 3.76); MS, m/z 365 ($M - 2$)⁺ (0.4), 364 (0.4), 351 (49), 323 (45), 322 (96), 320 (100), 308 (87). (+)-Epidihydroturkiyenine (**3**): ν_{max} (CHCl_3) 3550 cm^{-1} ; λ_{max} MeOH 226, 286, 313 nm ($\log \epsilon$ 4.44, 3.86, 3.79); MS, m/z 365 ($M - 2$)⁺ (0.3), 363 (0.3), 362 (1), 323 (46), 322 (100), 320 (47), 308 (96). NMR spectra for **2** and **3** are in CD_3CN .

(8) Pseudobenzylisoquinolines always incorporate three oxygenated substituents in the lower pendant aromatic ring and are possibly derived in the plant from the oxidation of protoberberinium salts. See: Murugesan, N.; Shamma, M. *Tetrahedron Lett.* **1979**, 4521. Shamma, M.; Guinaudeau, H. *Tetrahedron*, in press.

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Synthesis and DNA Binding and Photonicking Properties of Acridine Orange Linked by a Polymethylene Tether to (1,2-Diaminoethane)dichloroplatinum(II)

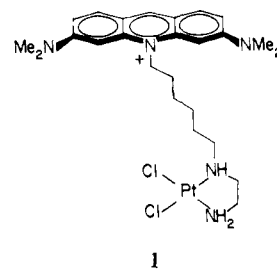
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There is much current interest in the binding of the antitumor drug *cis*-diamminedichloroplatinum(II) (*cis*-DDP) to its putative target in the cancer cell, DNA.¹ Both the position and mode of binding of *cis*-DDP to DNA can be altered by the presence of the intercalating dye ethidium bromide in the incubation medium.² Moreover, prior coordination of *cis*-DDP or $[\text{Pt}(\text{en})\text{Cl}_2]$ ($\text{en} = 1,2$ -diaminoethane) is known to affect the binding of intercalators to DNA.³ We were therefore interested to construct a molecule in which both an intercalating functionality and a diamine-coordinated $[\text{PtCl}_2]$ moiety are connected by an appropriate linker chain and to study its DNA binding and cleaving properties. There is precedence for compounds containing both intercalator and metal-binding functionalities in the naturally occurring antitumor antibiotic bleomycin,⁴ in a family of synthetic molecules designed as footprinting agents,⁵ and in certain metallointercalation reagents such as $[\text{Pt}(\text{terpy})\text{Cl}]\text{Cl}$.^{3a} Here we report the synthesis, characterization, DNA binding, and photoactivated DNA cleaving (nicking) properties of cation **1**, in which acridine orange is linked by a hexamethylene chain to (1,2-diaminoethane)dichloroplatinum(II).

Compound **1** was synthesized in overall 18% yield by the following nine-step procedure. The hydroxyl group of 6-chloro-1-



hydroxyhexane was protected with dihydropyran,⁶ following which the chloro group was converted to an iodo group by a Finkelstein reaction.⁷ Alkylation of acridine orange free base⁸ by refluxing in xylene with a trace of NaHCO_3 , deprotection of the alcohol,⁹

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